

Hemoglobin An overview and more

Prof. Mamoun Ahram Hematopoietic-lymphatic system

Resources

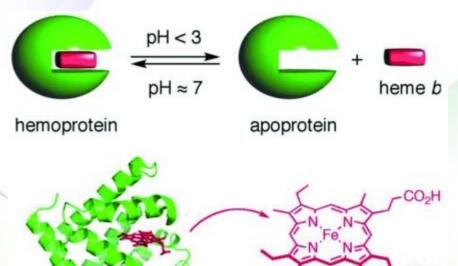


- This lecture
- Myoglobin/Hemoglobin O2 Binding and Allosteric Properties of Hemoglobin (http://home.sandiego.edu/~josephprovost/Chem331%20Lect%207_8%20Myo%20Hemoglobin.pdf)
- Lecture 3: Cooperative behaviour of hemoglobin (https://www.chem.uwec.edu/chem452 f12/pages/lecture materials/unit III/lecture-3/overheads/Chem452-lecture 3-part 1-overheads.pdf)

Hemoproteins

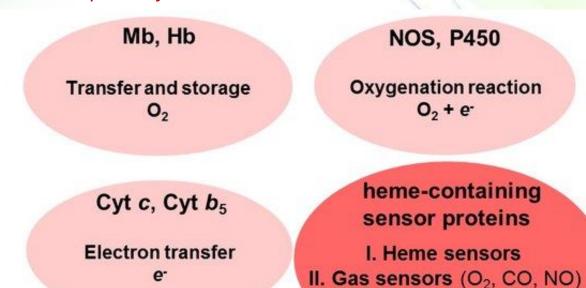


Many proteins have heme as a prosthetic group called hemoproteins.



native heme b (1)

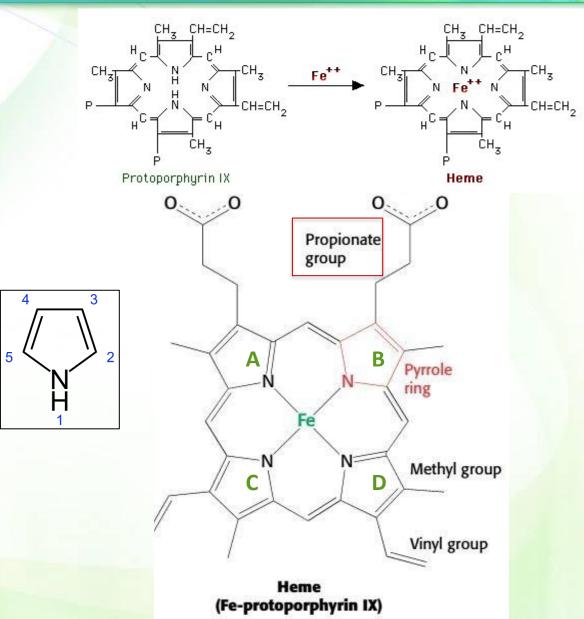
A prosthetic group is a tightly bound, specific non-polypeptide unit required for the biological function of some proteins. The prosthetic group may be organic (such as a vitamin, sugar, or lipid) or inorganic (such as a metal ion), but is not composed of amino acids.



Heme structure



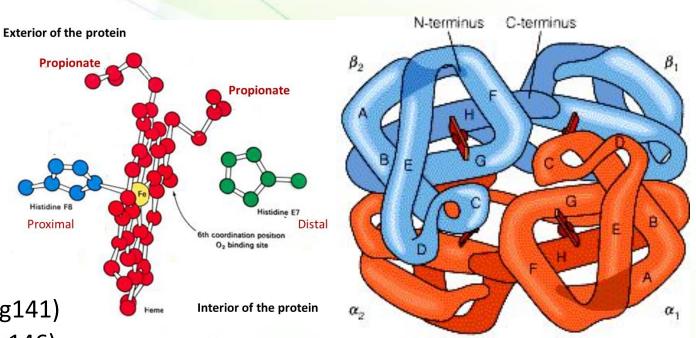
- It is a complex of protoporphyrin IX + iron (Fe²⁺).
- The porphyrin is planar and consists of four rings (designated A-D) called pyrrole rings.
- Each pyrrole can bind two substituents.
- Two rings have a propionate group each.
- Note: the molecule is hydrophobic.
- Fe has six coordinates of binding.

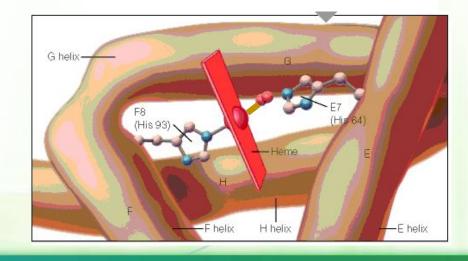


Structure of hemoglobin



- Hb is a globular protein.
 - Typical amino acid distribution
 - Positions of two histidine residues
 - Proximal and distal
- It is an allosteric protein.
 - Multiple subunits $(2\alpha + 2\beta)$
 - α polypeptide = 141 amino acids (Arg141)
 - β polypeptide = 146 amino acids (His146)
 - The first amino acid in both is valine.
 - Altered structure depending on bound molecules
 - Positive cooperativity towards oxygen
 - Regulated by allosteric effectors

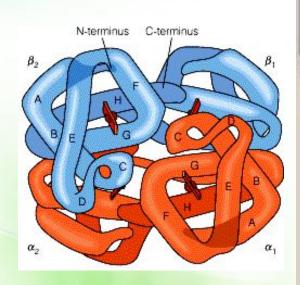


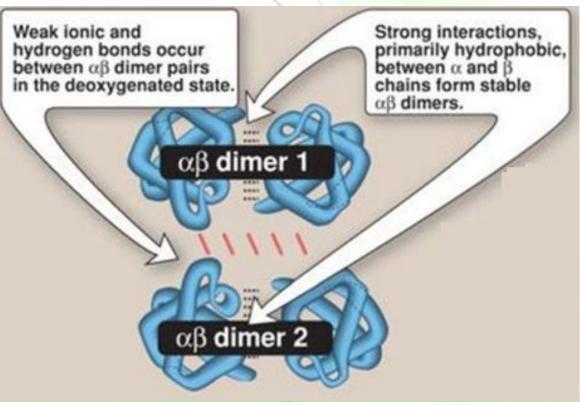


How are the subunits bound?



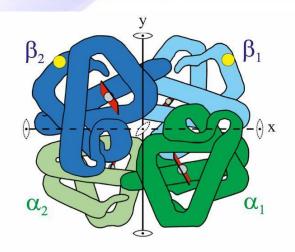
- A dimer of dimers (I made up this term)
 - (α-β)2
 - Note how they interact with each other.

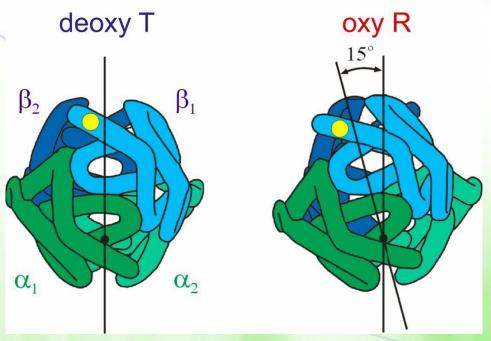


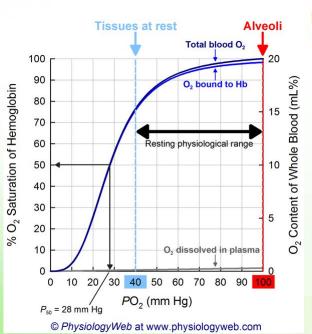


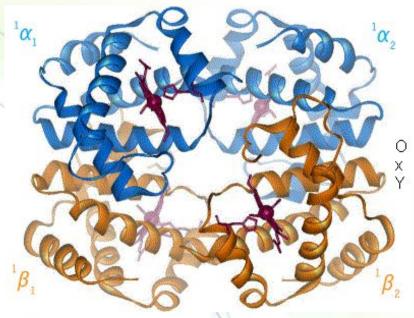
Structural change of hemoglobin







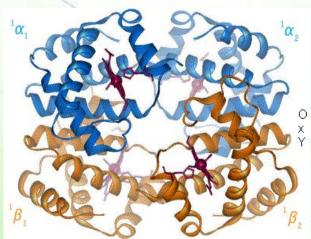


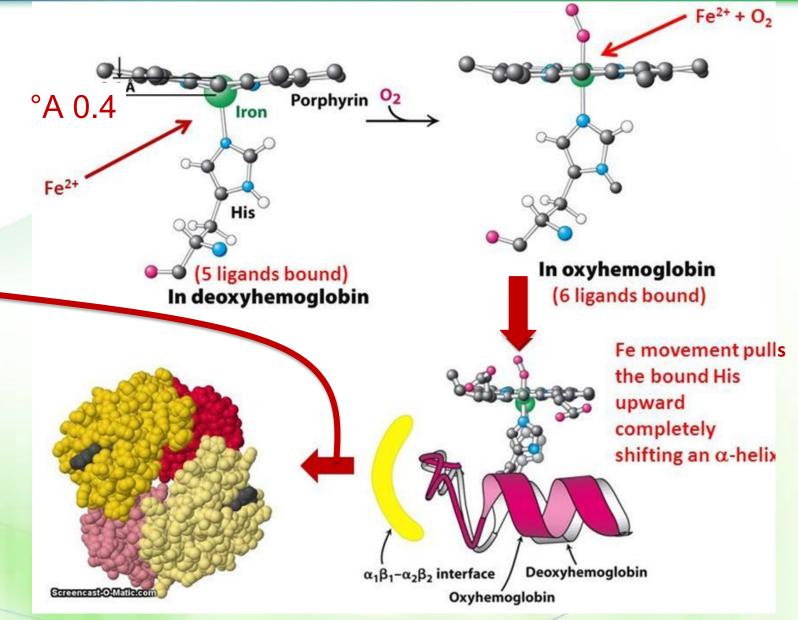


Structural amplification change



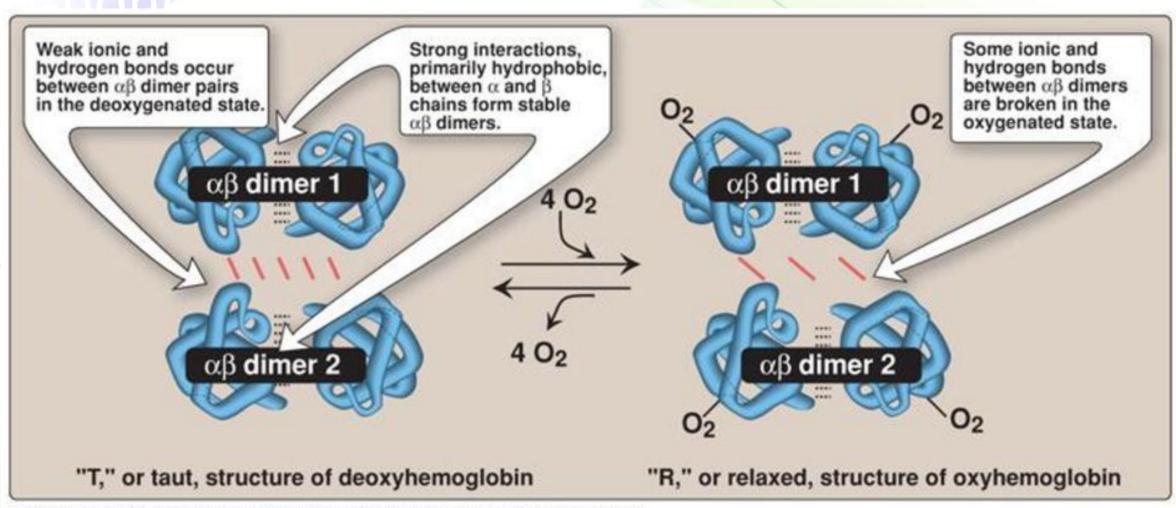
- Changes in tertiary structure of individual hemoglobin subunits
- Breakage of the electrostatic bonds at the other oxygen-free hemoglobin chains.





Broken electrostatic interactions and





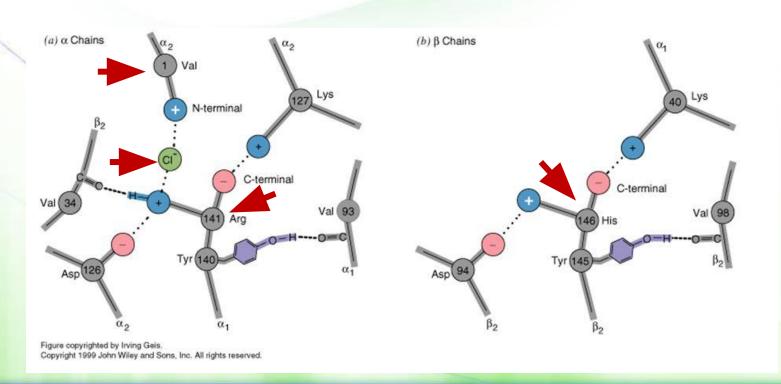
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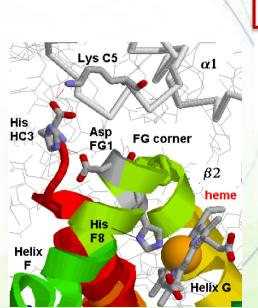
The broken interactions

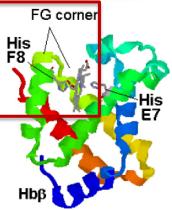


• Electrostatic interactions and hydrogen bonds (at the C-termini of the alpha and beta chains) that stabilize the T-form of hemoglobin are broken upon movement of the alpha-helix.

Note the groups, the protonation status, and the allosteric effectors



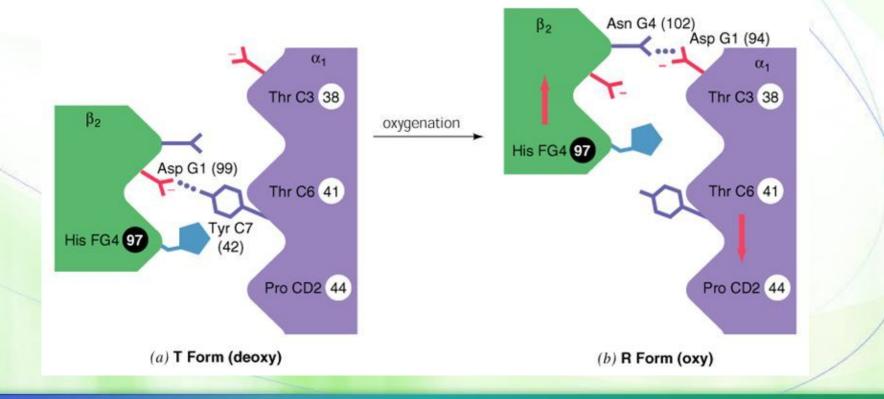




Reformation of hydrogen bonds

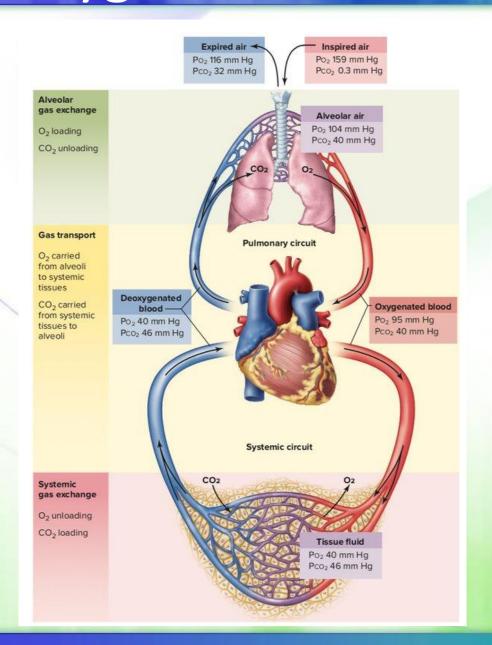


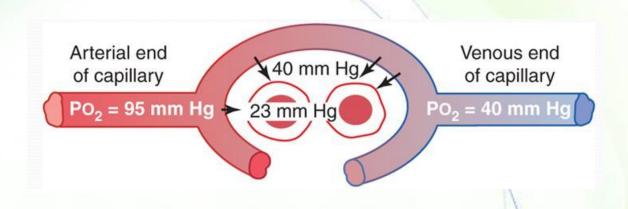
- T-state hemoglobin (deoxyhemoglobin) is stabilized by a hydrogen bond between Asp G1 (99) of β 2 with Thr C6 (41) of α 1.
- When O_2 binds, the $\alpha 1$ surface slides and a hydrogen bond is formed between Asn G4 (102) of β chain and Asp 1 (94) of α chain stabilizing the R form of hemoglobin.



Oxygen distribution in blood versus



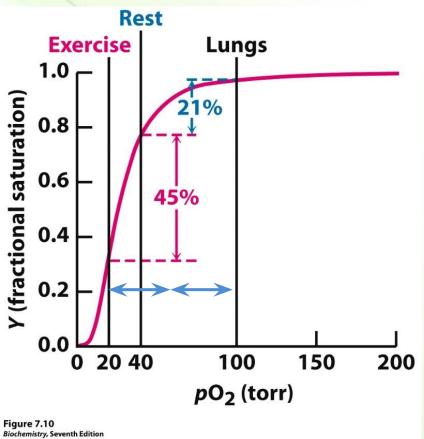




Oxygen saturation curve



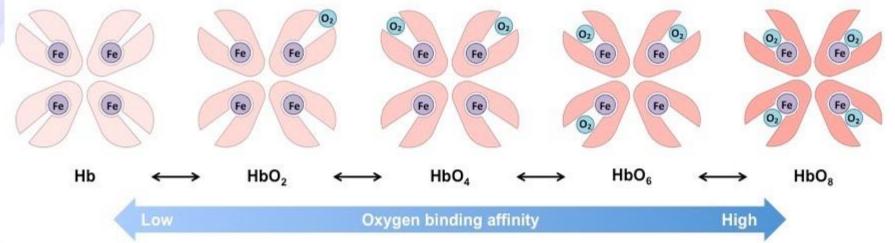
- The saturation curve of hemoglobin binding to O₂ has a sigmoidal shape.
 - It is allosteric.
- At 100 mm Hg, hemoglobin is 97% saturated (oxyhemoglobin).
- As the oxygen pressure falls, oxygen is released to the cells.
- Note: at high altitude (~5000 m), alveolar pO2 = 75 mmHg.



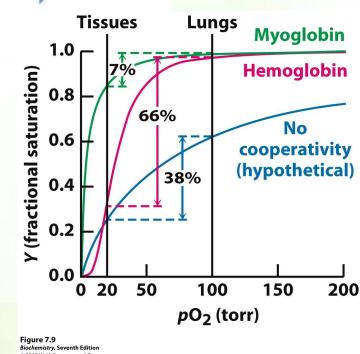
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Positive cooperativity

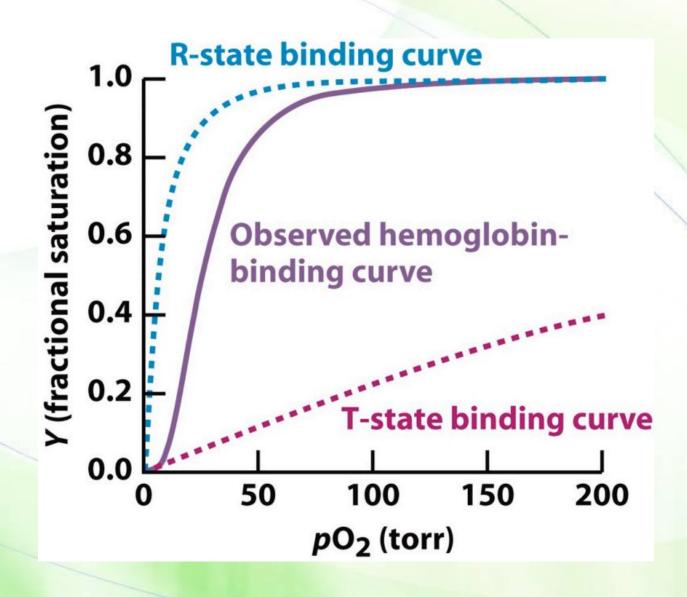




- Increasing ligand concentration drives the equilibrium between R and
 T toward the R state (positive cooperativity) sigmoidal curve
 - The effect of ligand concentration on the conformational equilibrium is a **homotropic effect (oxygen)**.
 - Other effector molecules that bind at sites distinct from the ligand binding site and thereby affect the R and T equilibrium in either direction are called **heterotropic effectors** (e.g. CO₂).







The Hill constant (coefficient)

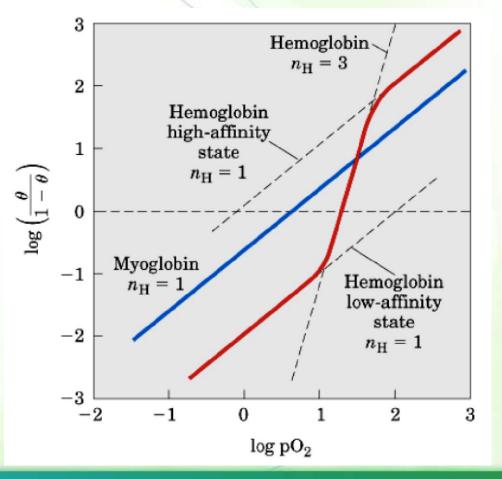


- The Hill plot is drawn based on an equation (you do not have to know it).
- n = Hill constant determined graphically by the hill plot
- n is the slope at the midpoint of binding of log (Y/1-Y) vs. log of pO₂.
 - if n = 1 then non cooperativity
 - if n < 1 then negative cooperativity
 - if n >1 then positive cooperativity
- The slope reflects the degree of cooperativity, not the number of binding sites.

$$\log \frac{Y}{1 - Y} = n \log pO \quad 2^{-n} \log P$$

$$Y \text{ or } \theta \text{ is the fraction of oxygen-bound Hb}$$

$$\rightarrow Y = mX + b \text{ (linear plot)}$$



Cooperativity models



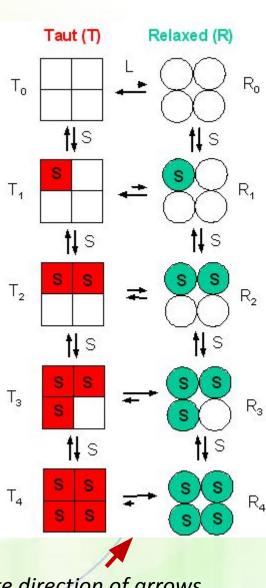
- Two models of cooperativity that could explain the observed data
 - Concerted model all subunits undergo the conformational change simultaneously
 - There are only two states, R and T.
 - Sequential model the subunits undergo the conformational change one at a time.
 - There are multiple states between full T and full R.

The concerted model (MWC



rodel

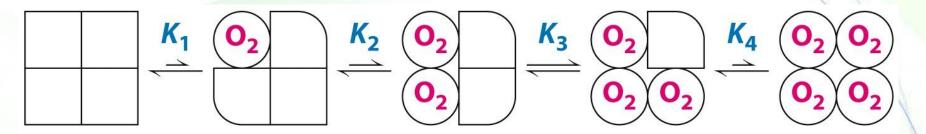
- The protein exists in two states in equilibrium: T (taut, tense) state with low affinity and R (relaxed) state with high affinity.
- Increasing occupancy increases the probability that a hemoglobin molecule will switch from T to R state.
- This allows unoccupied subunits to adopt the high affinity R-state.



The sequential, induced fit, or KNF model



 The subunits go through conformational changes independently of each other, but they make the other subunits more likely to change, by reducing the energy needed for subsequent subunits to undergo the same conformational change.



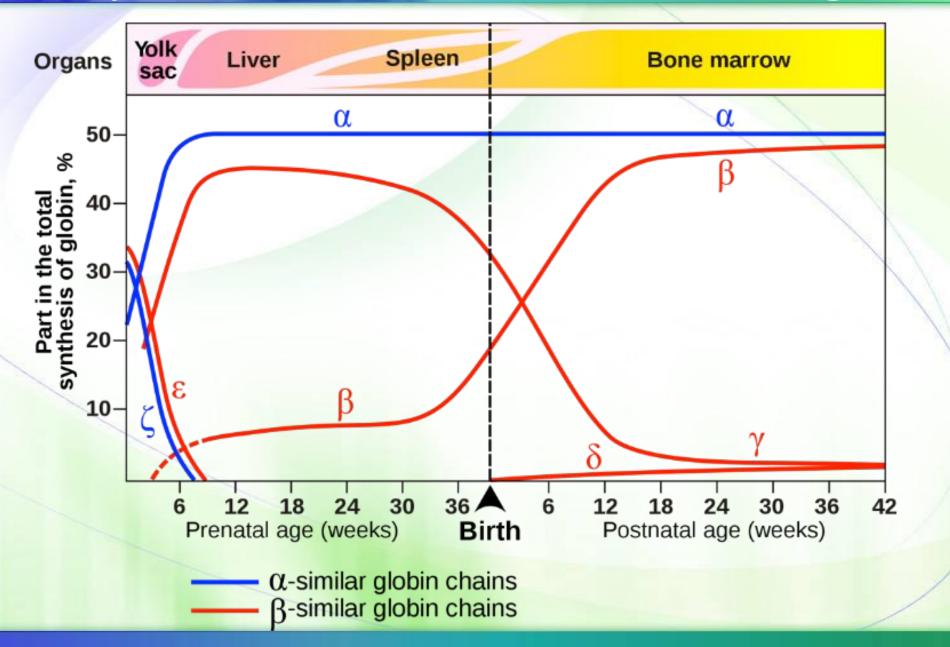
Which one is better? Both can explain the sigmoidal binding curve.



It is not only one hemoglobin

Developmental transition of hemoglobins





Alpha $\mathbf{A} \alpha$ Beta $\mathbf{B} \beta$ Gamma $\mathbf{\Gamma} \gamma$ Delta $\Delta \delta$ Epsilon $\mathbf{E} \varepsilon$ Zeta $\mathbf{Z} \zeta$

The embryonic stage

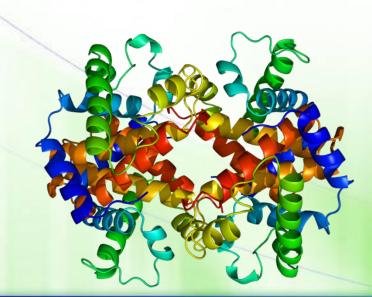


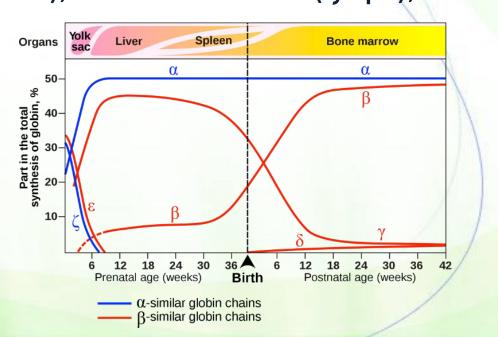
 Hemoglobin synthesis begins in the first few weeks of embryonic development within the yolk sac.

• The major hemoglobin (HbE Gower 1) is a tetramer composed of 2 zeta (ξ) chains and 2 epsilon (ϵ) chains

• Other forms exist: HbE Gower 2 (α 2 ϵ 2), HbE Portland 1 (ζ 2 γ 2), HbE

Portland 2 (ζ 2 β 2).

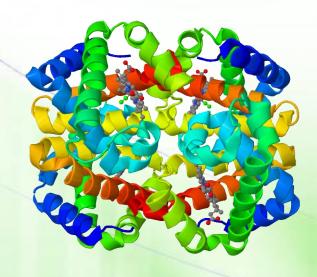


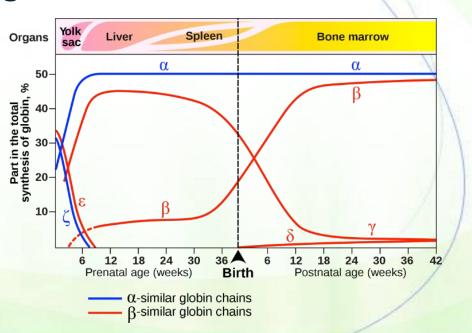


The fetal stage



- By 6-8 weeks of gestation, the expression of embryonic hemoglobin declines dramatically and fetal hemoglobin synthesis starts from the liver.
- Fetal hemoglobin consists of two α polypeptides and two gamma (γ) polypeptides ($\alpha 2\gamma 2$)
- The α polypeptides remain on throughout life.

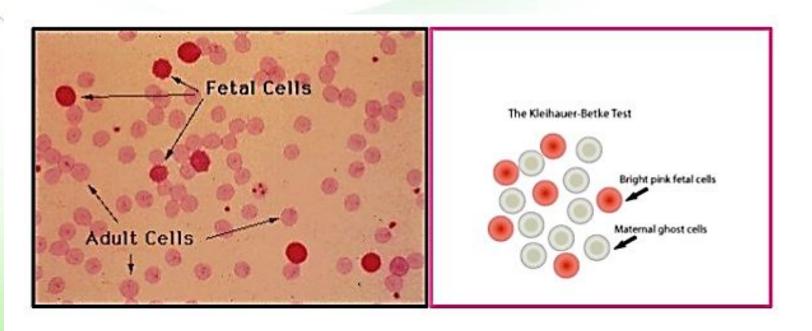


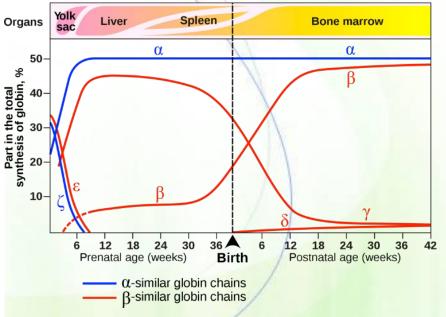


The adult stage



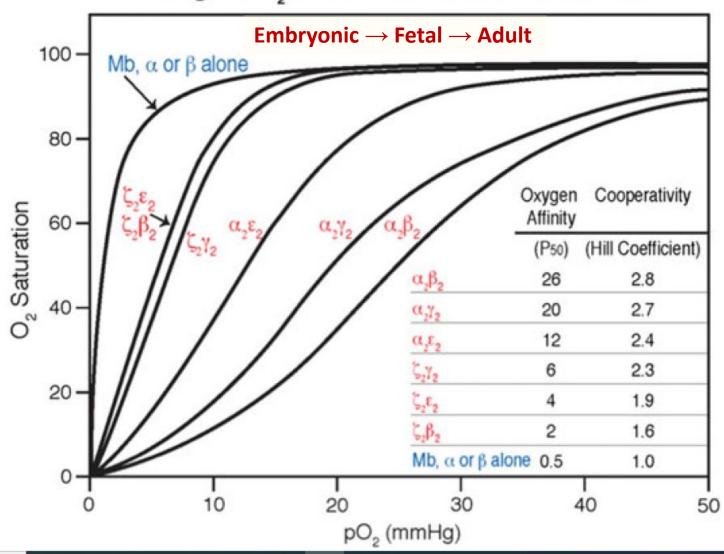
- Shortly before birth, there is a gradual switch to adult β-globin.
- Still, HbF makes up 60% of the hemoglobin at birth, but 1% of adults.
- At birth, synthesis of both γ and β chains occurs in the bone marrow.







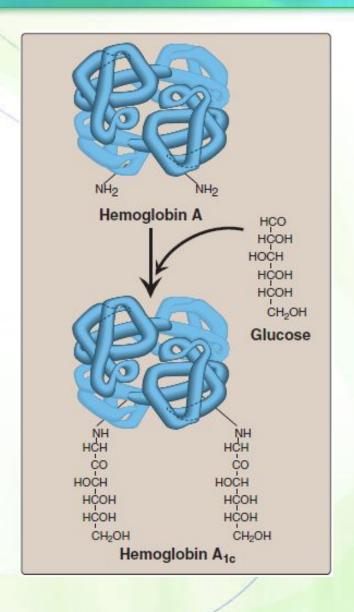




Adult hemoglobins



- The major hemoglobin is HbA1 (a tetramer of 2 α and 2 β chains).
 - A minor adult hemoglobin, HbA2, is a tetramer of 2 α chains and 2 delta (δ) chains.
- HbA1 can be glycosylated with a hexose and is designated as HbA1c.
 - The major form (HbA1c) has glucose molecules attached to valines of β chains.
 - HbA1c is present at higher levels in patients with diabetes mellitus.



Advantages of HbA1c testing



- <u>Blood fasting glucose</u> level is the concentration of glucose in blood at a single point in time when fasting for a few hours.
- <u>HbA1c</u> level provides <u>a longer-term trend</u>, similar to an average, of how high blood sugar levels have been over a period of time (2-3 months).
- HbA1c can be expressed as a percentage (DCCT unit, used in the US) or as a value in mmol/mol (IFCC unit).

Table



BLOOD GLUCOSE		STATUS	HbA1c	
mmol/L	mg/dL		%	mmol/mol
5.4	97	Normal	5	31
7.0	126		6	42
8.6	155	Pre-Diabetes	7	53
10.2	184	Diabetes	8	64
11.8	212	Diabetes	9	75
13.4	241		10	86
14.9	268	Diabetes	11	97
16.5	297		12	108

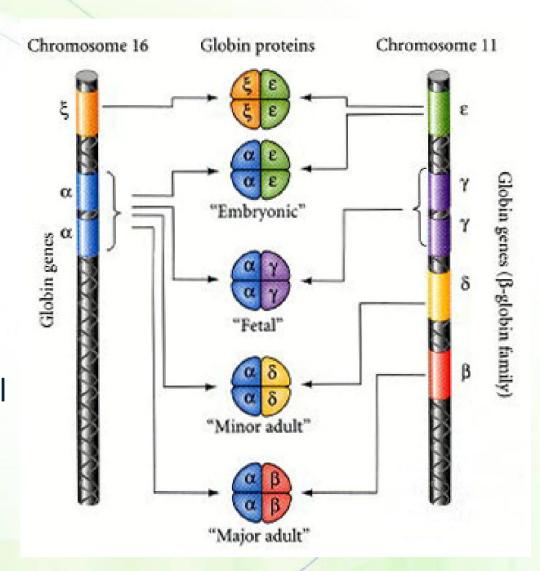


Genetics of globin synthesis

The genes



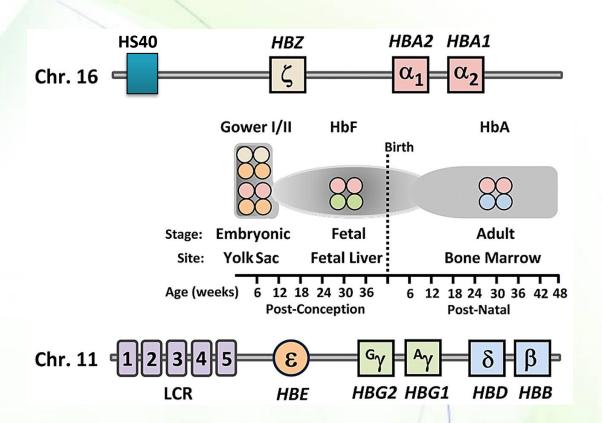
- The α gene cluster contains three genes: two α genes (α 1 and α 2), and ζ (zeta) gene.
- The β gene cluster contains five genes: β gene, ϵ (epsilon) gene, two γ (gamma) genes, and δ (delta) gene.
- The gene order parallels the order of expression.
- Genetic switching is controlled by a transcription factor-dependent developmental clock, independent of the environment.
- Premature newborns follow their gestational age.



Locus structure



- Each gene has its promoter and regulatory sequences (activators, silencers).
- The α gene cluster is controlled by the HS40 region.
- The β-globin cluster is controlled by a master enhancer called locus control region (LCR).

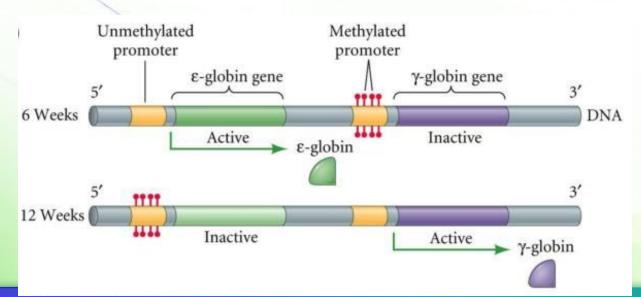


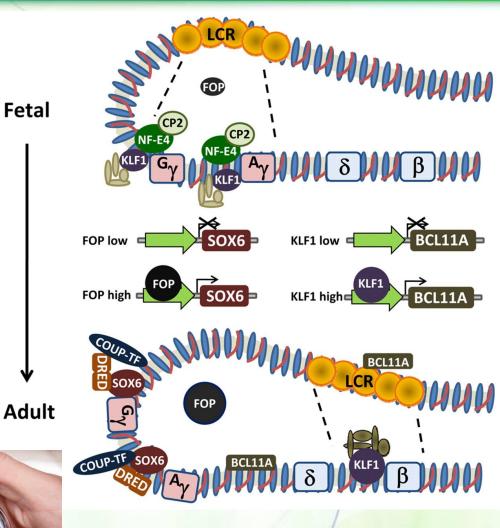
The mechanism of regulation



• The mechanism requires *timed* expression of regulatory transcription factors for each gene, epigenetic regulation (e.g., acetylation, methylation), chromatin looping, and non-coding RNA (e.g., long non-coding RNA, microRNA, etc.).

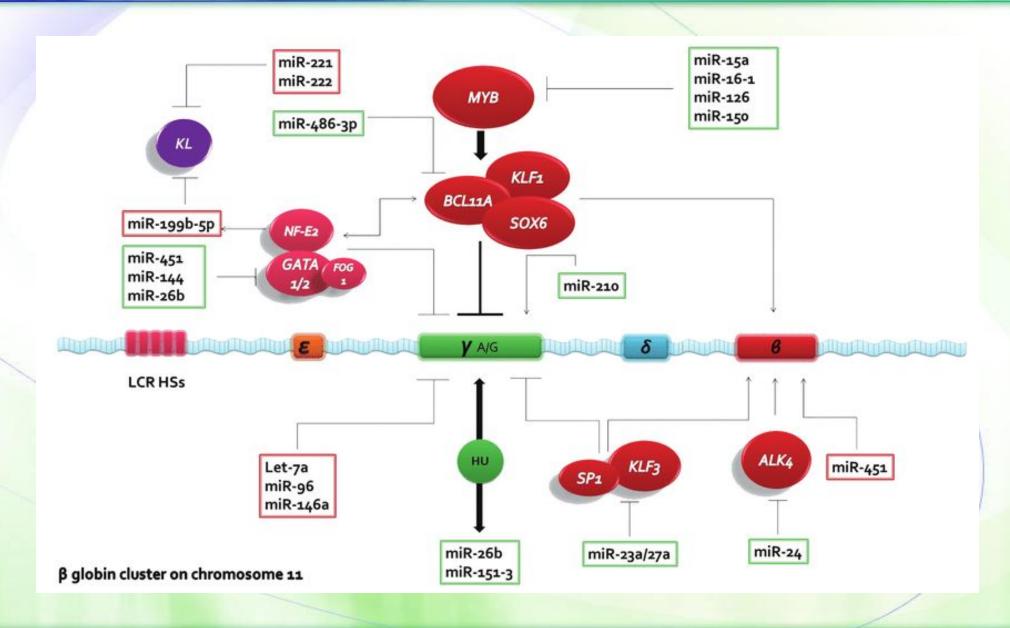
Note: treatment!!





For entertainment only!!







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Activation of γ-globin expression by hypoxia-inducible factor 1a

Ruopeng Feng, Thiyagaraj Mayuranathan, Peng Huang, Phillip A. Doerfler, Yichao Li, Yu Yao, Jingjing

Zhang, Lance E. Palmer, Kalin Mayberry, Georgios E. Christakopoulos, Peng Xu, Chunliang Li, Yong Cheng,

Gerd A. Blobel, M. Celeste Simon & Mitchell J. Weiss

☐



Abstract

Around birth, globin expression in human red blood cells (RBCs) shifts from γ-globin to βglobin, which results in fetal haemoglobin (HbF, $\alpha_2 \gamma_2$) being gradually replaced by adult haemoglobin (HbA, $\alpha_2\beta_2$)¹. This process has motivated the development of innovative approaches to treat sickle cell disease and β-thalassaemia by increasing HbF levels in postnatal RBCs². Here we provide therapeutically relevant insights into globin gene switching obtained through a CRISPR-Cas9 screen for ubiquitin-proteasome components that regulate HbF expression. In RBC precursors, depletion of the von Hippel-Lindau (VHL) E3 ubiquitin ligase stabilized its ubiquitination target, hypoxia-inducible factor 1α (HIF1 α)^{3,4}, to induce γ-globin gene transcription. Mechanistically, HIF1α-HIF1β heterodimers bound cognate DNA elements in BGLT3, a long noncoding RNA gene located 2.7 kb downstream of the tandem γ-globin genes *HBG1* and *HBG2*. This was followed by the recruitment of transcriptional activators, chromatin opening and increased long-range interactions between the y-globin genes and their upstream enhancer. Similar induction of HbF occurred with hypoxia or with inhibition of prolyl hydroxylase domain enzymes that target HIF1α for ubiquitination by the VHL E3 ubiquitin ligase. Our findings link globin gene regulation with canonical hypoxia adaptation, provide a mechanism for HbF induction during stress erythropoiesis and suggest a new therapeutic approach for β-haemoglobinopathies.